



# A rapid colorimetric detection method of trace Cr (VI) based on the redox etching of Ag<sub>core</sub>-Au<sub>shell</sub> nanoparticles at room temperature

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## ABSTRACT

A rapid colorimetric detection method of trace Cr (VI) in aqueous solutions has been developed based on non-aggregated Ag<sub>core</sub>-Au<sub>shell</sub> nanoparticles. It is based on the fact that Cr (VI) redox etches Ag<sub>core</sub>-Au<sub>shell</sub> nanoparticles at the present of bromide ions of hexadecyl trimethyl ammonium bromide (CTAB). The etching process of Ag<sub>core</sub>-Au<sub>shell</sub> nanoparticles would lead to a blue shift in the surface plasmon resonance (SPR) absorption peak as the size of Ag<sub>core</sub>-Au<sub>shell</sub> nanoparticles decreased. This colorimetric strategy based on size and component dependence of core-shell nanoparticles during the etching process provided a highly sensitive and selective detection method toward Cr (VI). Compared with other detection methods, this method provided a wide linear detection range from  $1 \times 10^{-8}$  M to  $8 \times 10^{-6}$  M over one order of magnitude, also has some practical capability. The cost-effective probe in this colorimetric method allowed rapid and sensitive detection of trace Cr (VI) ions as low as  $1.0 \times 10^{-7}$  M based on the observations by the naked eyes and  $1.0 \times 10^{-8}$  M based on the measurements of UV-vis spectra in aqueous solutions at room temperature. According to the preliminary data, it indicated that the current method showed very promising practical applicability for the determination of Cr (VI) in real environmental samples.

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## 1. Introduction

In natural environment, chromium is generally in two most stable states of Cr (III) and Cr (VI), but their toxicities and mobilities differ significantly. On one hand, it is well known that Cr (III) participates in the metabolism of sugar and adipose in human beings and its deficiency may induce hyperglycemia, fat metabolism disorders, atherosclerosis, and coronary heart disease so on. [1,2] Moreover, chromium (III) is one of the essential trace elements in the human body and some chromium (III) compounds such as chromium (III) picolinate have been employed as supplemental or alternative medication for diabetes. [3] On the other hand, Cr (VI) is known to be highly soluble and toxic with carcinogenic effects, which is hazardous to human beings. [4] Therefore, much more attention has been attracted to the detection of Cr (VI) in a number of industrial processes such as chrome plating, dye and pigment fabrication, leather tanning, and wood preserving in recent years. Various sensor systems have been reported before.[5–13] Recently, a system performed by directed assembly of gold nanoparticles using DNA as a colorimetric sensor for the detection of metal ions has made a remarkable progress. [14,15] However, most of these systems have either limitations with aspect to sensitivity, simplicity or the need

of surface modification for substantial selectivity. Also, a non-aggregation based label-free colorimetric sensor for the detection of Cr (VI) has been developed based on the redox etching of gold nanorods, [16a] but the method for preparation of gold nanorods is complicated, time-consuming, need of heating, and hard to control the aspect ratio of gold nanorods. Moreover, the detection of Cr (VI) based on the redox etching of gold nanorods is not suitable at room temperature. Above all, each of these strategies has advantages and disadvantages and no single method has gained supremacy.

In this report, a rapid colorimetric method for the detection of trace Cr (VI) in aqueous solutions based on label-free and non-aggregation Ag<sub>core</sub>-Au<sub>shell</sub> nanoparticles has been developed at room temperature. This rapid and simple method is based on the fact that Cr (VI) redox can etch Ag<sub>core</sub>-Au<sub>shell</sub> nanoparticles when there are bromide ions in hexadecyl trimethyl ammonium bromide (CTAB). The difference in the thickness of gold shell nanostructures caused a change of the surface plasmon resonance (SPR) absorption in the UV-vis spectra. The regular changes of the SPR absorption in the UV-vis spectra were used for quantitative analysis in the detection of Cr (VI). It was found that this colorimetric assay method for the detection limit of Cr (VI) concentration was as low as  $1.0 \times 10^{-7}$  M based on the observations by the naked eyes and  $1.0 \times 10^{-8}$  M based on the measurements of UV-vis spectra. The linear relationship of the quantitative analysis for Cr (VI) concentrations ranged from  $8.0 \times 10^{-6}$  to  $1.0 \times 10^{-8}$  M. This work exhibits very high sensitivity and excellent selectivity in the detection of Cr (VI) along with low

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detection limit in aqueous solutions at room temperature. A real sample was tested to prove our method with the practicability for the environmental samples in aqueous solutions.

## 2. Experimental section

### 2.1. Materials and characterization

$\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$  (> 99.0%),  $\text{AgNO}_3$  ( $\geq 99.8\%$ ),  $\text{NaBH}_4$  ( $\geq 96.0\%$ ), trisodium citrate dihydrate ( $\text{C}_6\text{H}_5\text{Na}_3\text{O}_7 \cdot 2\text{H}_2\text{O}$ ,  $\geq 99.0\%$ ),  $\text{NH}_2\text{OH} \cdot \text{HCl}$  ( $\geq 98.5\%$ ), hexadecyl trimethyl ammonium bromide (CTAB, 99%), and other reagents (analytical-reagent grade) were purchased from Sinopharm Chemical Reagent Co. Ltd (Shanghai, China) without further purification as received. Transmission electron microscopy (TEM) was performed using a Tecnai F20 instrument operated at 200 kV. UV–vis spectroscopy was performed using a Lambda 950 instrument and inductively coupled plasma atomic emission spectroscopy (ICP-AES) was performed with an Optima 2100 instrument from Perkin Elmer. Nanopure deionized and double distilled water (18.2 M $\Omega$ ) was used for all experiments.

### 2.2. Synthesis of $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$ nanoparticles

$\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles were prepared according to the seed growth method of Cui et al. in a previous report with modification [17]. A batch of 100 mL solution with a final concentration of 0.2 mM  $\text{AgNO}_3$  and 0.5 mM  $\text{C}_6\text{H}_5\text{Na}_3\text{O}_7 \cdot 2\text{H}_2\text{O}$  in water was prepared. While stirring vigorously, 1.0 mL of 100 mM  $\text{NaBH}_4$  was added drop by drop into the mixture and the solution was kept at room temperature for 2 h. After that, the core-shell nanoparticles with varying molar fractions of Au were prepared by the following method:  $x$  mL of 62.5 mM  $\text{NH}_2\text{OH} \cdot \text{HCl}$  and  $x$  mL of 5 mM  $\text{HAuCl}_4$  were added dropwise (ca. 2 mL/min) by two separated pipettes into the silver colloid solutions (where  $x$  is the concentration of Au desired in the final product). Stirring was continued for another 45 minutes. The obtained  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles solution was centrifugated at 10,000 rpm for 15 min and then resuspended in 0.1 M CTAB with half of the original volume for use according to previous methods [18,19].

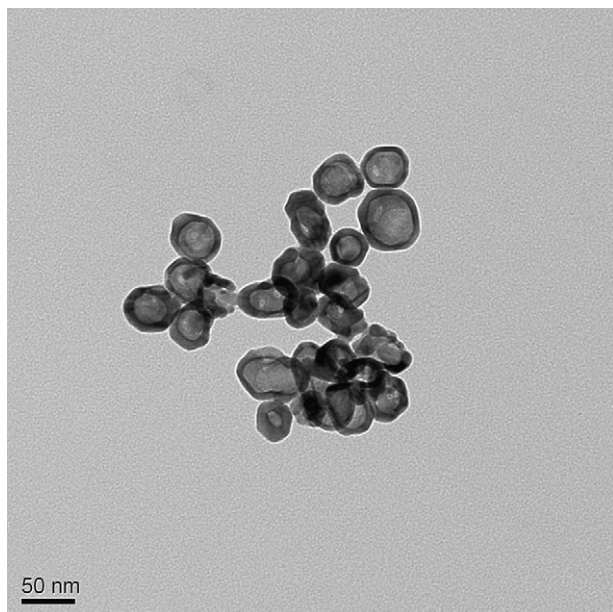


Fig. 1. TEM images of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles. The molar ratio of Au/Ag was 1.25.

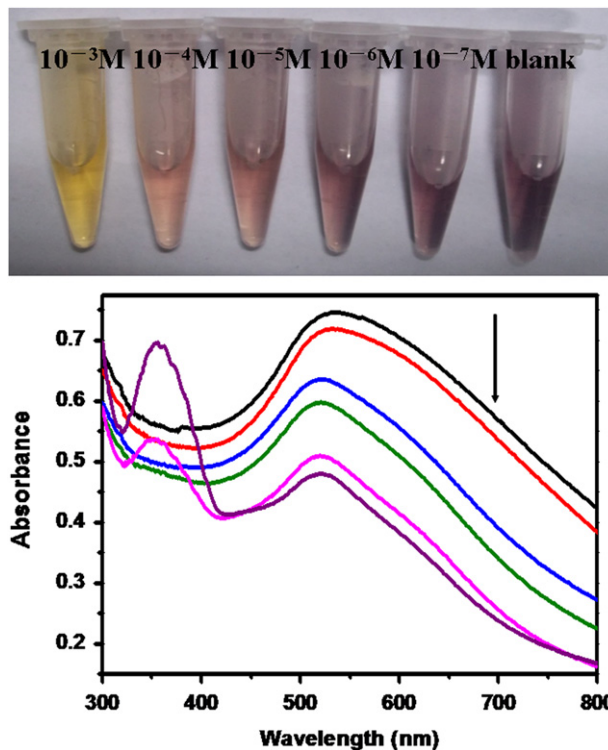


Fig. 2. (Top) Photographic images and (Bottom) surface plasmon resonance absorption change of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles in the presence of different concentrations of Cr (VI). (1) 0 M, (2)  $10^{-7}$  M, (3)  $10^{-6}$  M, (4)  $10^{-5}$  M, (5)  $10^{-4}$  M, (6)  $10^{-3}$  M (0.2 M CTAB; pH=2.00).

According to transmission electron microscopy data as shown in Fig. 1,  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles with the molar ratios of Au to Ag (1.25) were prepared.

### 2.3. $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$ nanoparticles solution for the detection of Cr (VI)

For the detection of Cr (VI) by using the  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles solution, different concentrations of Cr (VI) in aqueous solutions were added separately to a 2.0 mL of solution containing  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles, and adjusted the pH value of the mixtures to 2.0. As shown in Fig. 2, the mixtures were maintained at room temperature for only several minutes, and then the color changed from violet to yellow quickly.

## 3. Results and discussion

### 3.1. Sensing strategy

To understand the role that Cr (VI) plays in the redox etching of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles, the SPR absorption of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles was monitored under conditions of varying Cr (VI) concentrations shown in Fig. 2. The initial  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles exhibited the SPR absorption peak located at 535 nm and the SPR absorption peak gradually blue shifted and the absorbance decreased as the concentration of Cr (VI) increased. Moreover, a new SPR absorption peak was observed about 350–380 nm when the concentration of Cr (VI) reached to  $10^{-4}$  M. This is the evidence of silver nanoparticles after the redox etching of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles. Before Li et al. had also investigated the change of the absorption properties/selectivity of silver nanoparticles after complete etching/leaching of Au from the surface of Ag/Au nanoparticles by thiosulfate ( $\text{S}_2\text{O}_3^{2-}$ ), it proves the data in our experimental.[16b] Besides the color of

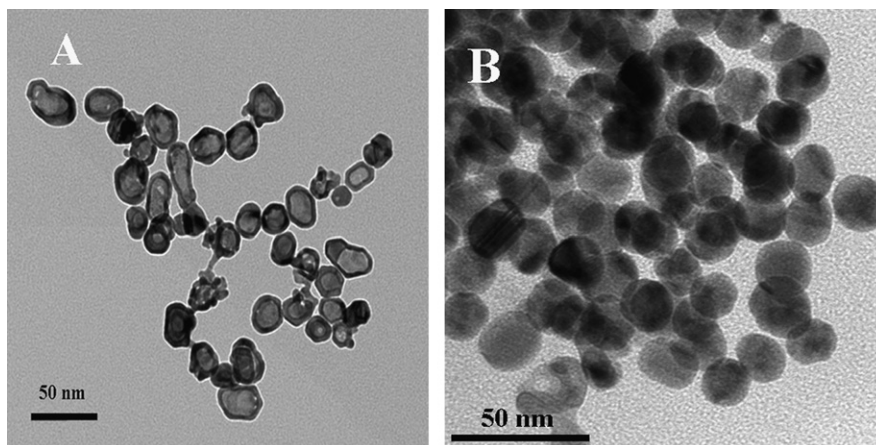
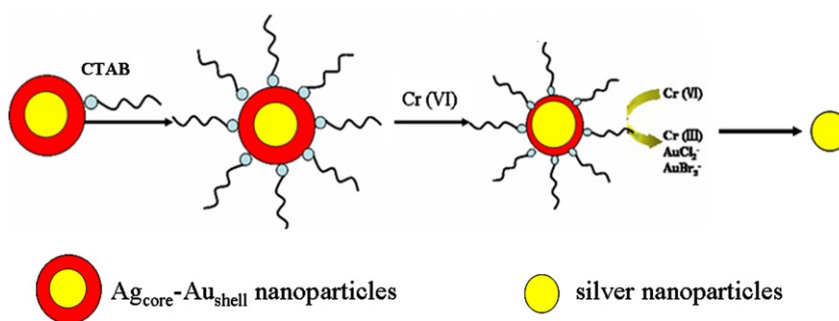


Fig. 3. TEM images of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles in the presence of different concentrations of Cr (VI). (A) 0 M, (B)  $10^{-3}$  M (0.2 M CTAB; pH=2.00).



Scheme 1. Schematic illustration on the redox etching mechanism of the  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles induced by Cr (VI).

$\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles solutions distinctly changed from violet to yellow. Taking into account of these observations and with regards to the investigations, it was shown that the blue-shift was due to the decreasing of particle size of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles, which agreed very well with the TEM images shown in Fig. 3.

Scheme 1 illustrated on a redox etching mechanism of the  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles induced by Cr (VI), which is employed in this method. A blue shift of the SPR wavelength is due to the decreasing of the size of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles, when the  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles reacted with Cr (VI) in aqueous solutions. The standard electron potential of Au (I)/Au (0) and Ag (I)/Ag (0) is 1.691 eV and 0.7966 eV, respectively. The electron potential of Au (I)/Au (0) and Ag (I)/Ag (0) decreases when the concentrations of  $\text{Br}^-$  of CTAB and  $\text{Cl}^-$  of HCl increase, and they act as ligands of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles ( $\text{AuBr}_2^- + e^- \rightarrow \text{Au} + 2\text{Br}^-$ ,  $E = 0.959$  eV, [20]  $\text{AuCl}_2^- + e^- \rightarrow \text{Au} + 2\text{Cl}^-$ ,  $E = 1.15$  eV, [21]  $\text{AgBr} + e^- \rightarrow \text{Ag} + \text{Br}^-$ ,  $E = 0.07133$  eV,  $\text{AgCl} + e^- \rightarrow \text{Ag} + \text{Cl}^-$ ,  $E = 0.22233$  eV). The standard electron potential of Cr (VI)/Cr (III) (1.33 eV) is higher than that of Au (I)/Au (0) and Ag (I)/Ag (0) in the presence of  $\text{Br}^-$  and  $\text{Cl}^-$  ions, which enables Cr (VI) to oxidize  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles. The redox etching process induced by Cr (VI) causes a significant decrease for the thickness of gold shell. When the concentration of Cr (VI) reached a higher level, upon further oxidation, the size of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles decreased, and the core-shell nanoparticles converted into silver nanospheres along with eventually completely oxidization, which exhibited different color to change from violet to yellow.

### 3.2. Optimization of the conditions for Cr (VI) measurement

In order to optimize the conditions for the determination of Cr (VI), a number of parameters that influence on the sensitivity,

selectivity, accuracy and stability were investigated in a univariate approach.

#### (1) Effects of the thickness and size of $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$ nanoparticles

The thickness and concentration of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles in the detection system were examined in order to obtain a wide linear range and high sensitivity. Fig. S1 showed the TEM images of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles with different thickness. Moreover, Au/Ag composite nanoparticles were generated with the increased amount of  $\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$ , and it would affect the detection limit of Cr (VI) (shown in Fig. S2). From these pictures, sample D (shown in Fig. S2) showed a clearly core-shell structure of Au and Ag with the molar ratio of 1.25. The lower the concentration of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles, the redox etching rate of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles induced by Cr (VI) increased and blue shift accordingly enhanced, with a favorable sensitivity but narrow linear range compared with high concentration of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles. In contrast, higher concentration of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles expanded the linear range but limited the sensitivity. In consideration of both linear range and sensitivity,  $3.6 \times 10^{-4}$  M of silver atoms in  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles were chosen. In the first step of the reaction, excessive  $\text{NaBH}_4$  was added, therefore,  $\text{AgNO}_3$  was completely transformed into silver nanoparticles. As shown in Fig. 1,  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles were well prepared in the TEM image. Given all that, the concentration of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticle solution can be calculated from the amount of silver nitrate.

#### (2) Effects of pH and CTAB

As we all know that the redox etching rate of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles enhanced as the pH value decreased [16a], which ascribed to the electron potential and oxidation capability of Cr (VI) increased. To support this point, simple calculations were conducted to roughly estimate the electron potentials of



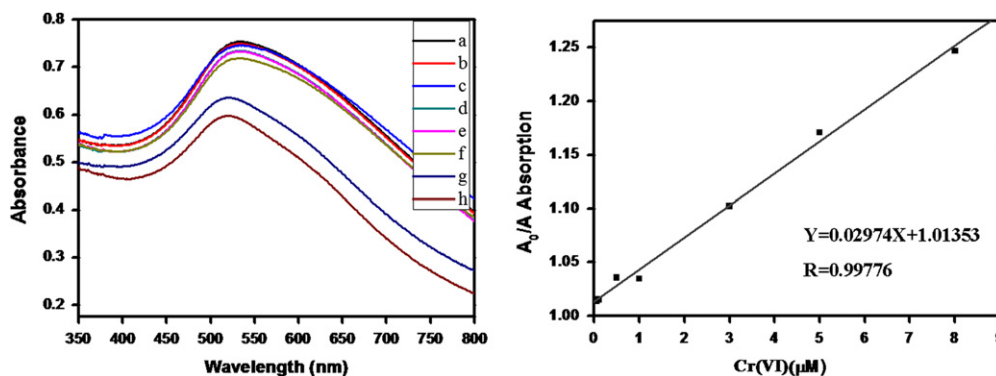


Fig. 5. (A) Surface plasmon resonance absorption changes of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles in the presence of different concentrations of Cr (VI). (a) 0  $\mu\text{M}$ , (b) 0.01  $\mu\text{M}$ , (c) 0.05  $\mu\text{M}$ , (d) 0.1  $\mu\text{M}$ , (e) 0.5  $\mu\text{M}$ , (f) 1  $\mu\text{M}$ , (g) 5  $\mu\text{M}$ , (h) 8  $\mu\text{M}$ . (B) The calibration curve for the detection of Cr (VI) by  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles. The ratio ( $A_0/A$ ) was plotted against different concentrations of Cr (VI). Where  $A_0$  is the UV absorption of the  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles and  $A$  is the UV absorption of the  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles in the presence of different concentrations of Cr (VI) at 526 nm.

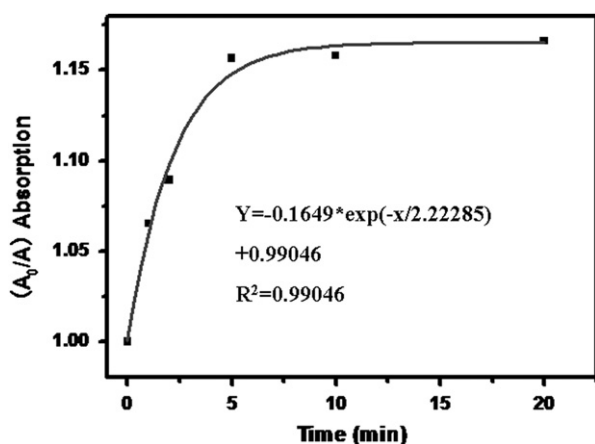


Fig. 6. The redox etching time of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles in the presence of Cr (VI) (2  $\mu\text{M}$ ) at room temperature.

(from Ningbo Environment Monitoring Center, Ningbo, China) was determined by applying this assay method. First, the pH of real water samples was adjusted to 9.0 with a certain amount of ammonia to precipitate  $\text{Cr}^{3+}$  and stirred for 30 min at room temperature. Then the pH values of the treated water samples were adjusted to 2.0 with HCl before use. A calibration curve of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles SPR shifts in the presence of different concentrations of Cr (VI) was prepared (see Fig. 5B). The concentration of Cr (VI) in the real water samples (from Ningbo Environment Monitoring Center, Ningbo, China) was from 1.60  $\mu\text{M}$  to 1.75  $\mu\text{M}$  based on the detection results, which was generally in good agreement with the detection result of Cr (VI) (1.63  $\mu\text{M}$ ) by the ICP-AES-based method. Thus, the result demonstrated that the currently designed colorimetric method is applicable for the detection of Cr (VI) in real environmental water samples.

## 5. Conclusions

A highly sensitive and selective colorimetric sensor is designed for the rapid detection of Cr (VI) at room temperature based on the redox etching process of non-aggregated  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles which is induced by Cr (VI). Cr (VI) was the only metal ion which brought a color change of the sample solution from violet to yellow and decreased the absorption of the SPR with  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles. The optimal pH value of  $\text{Ag}_{\text{core}}\text{-Au}_{\text{shell}}$  nanoparticles solution was determined to be 2.0.

In comparison with aggregation based colorimetric sensor based on cross-linking or electrostatic absorption, the proposed non-aggregation colorimetric sensor shows high selective and sensitive response towards Cr (VI) without surface modification. Some tests of the Cr (VI) concentration with the real water sample were done and the results were consistent with the ICP-AES-based method. In principle, this method could provide a promising sensor to on-line rapidly detecting Cr (VI) in real water samples.

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## Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at <http://dx.doi.org/10.1016/j.talanta.2012.09.009>.

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